

## Allura red

### Toxicological Data on the Unburnt Ingredient

[+ve, positive; -ve, negative; ?, equivocal; with, with metabolic activation; without, without metabolic activation]

#### In vivo

Species	Test conditions	Endpoint	Results	Reference
Mouse, CD-1 ICR strain (groups of 4 females)	Comet assay. Animals given a single dose of 2 g/kg bw by gavage on day 11 of pregnancy, and assessed for DNA damage in the brain, lung, liver, kidney, glandular stomach, colon, urinary bladder and embryo cells 3, 6 and 24 hr after treatment.	DNA damage	+ve  DNA damage was observed in the colon after 3 hr.	Tsuda et al. 2001
Mouse, CD-1 ICR strain (groups of 4 males)	Comet assay. Animals given a single dose of 0, 1, 10, 100, 1000 or 2000 mg/kg bw by gavage and assessed for DNA damage in the brain, lung, liver, kidney, glandular stomach, colon, urinary bladder and bone marrow	DNA damage	+ve  Dose-related DNA damage was observed in the colon and stomach after 3 hr. The colon was the only organ affected after 24 hr.	Tsuda et al. 2001

	3 (all doses), 6 and 24 hr (2000 mg/kg bw only) after treatment.			
Mouse, ddY strain (groups of 4 males)	Comet assay. Animals given a single dose of 0, 1, 10, 100, 1000 or 2000 mg/kg bw by gavage and assessed for DNA damage in the brain, lung, liver, kidney, glandular stomach, colon, urinary bladder and bone marrow 3 (all doses) and 24 hr (2000 mg/kg bw only) after treatment.	DNA damage	+ve  Dose-related DNA damage was observed in the colon and stomach after 3 hr. The colon was the only organ affected after 24 hr.	Sasaki et al. 2002a & b; Kawaguchi et al. 2001
Mouse (males, numbers unknown)	Animals given 4000 and 20,000 ppm (up to about 2.6 g/kg bw/day) in the diet for 8 weeks. Each male was then mated with 2 untreated females and the males from the F <sub>1</sub> generation were subsequently mated with untreated females to	Heritable translocation	-ve	Jorgenson et al. 1978

	<p>assess the induction of heritable translocation.</p> <p>No further details of this unpublished study were available from the expert review.</p>			
<i>Drosophila melanogaster</i>	<p>Fed 'at the LD<sub>50</sub> dose for 24 days' and examined for recessive lethal effects (chromosome loss, mutation at specific loci, and sex-linked recessive lethal damage (SLRL) and chromosomal translocation).</p> <p>No further details of this unpublished study were available from the expert review.</p>	Chromosome damage, mutation and SLRL mutation	<p>-ve</p> <p>SLRL was not increased when compared with the study controls but was higher when compared with controls combined from three studies. Concluded to be negative by the EPA (Lee et al. 1983).</p>	Anon, 1977a & 1978

#### In vitro

Test system	Test conditions	Endpoint	Activation status	Results	Reference
<i>Salmonella typhimurium</i> , strains TA97, TA98, TA100, TA1535	Ames test. Tested in a preincubation assay at concentrations of up to 10 mg/plate.	Mutation	With and without rat and hamster liver S9	<p>-ve</p> <p>A good quality study.</p>	NTP

<i>Salmonella typhimurium</i> , strains TA98, TA100, TA1535, TA1537, TA1538	Ames test. Tested at concentrations of up to 1 mg/plate.	Mutation	With and without	-ve	Brown et al. 1978
<i>Salmonella typhimurium</i> , strains TA98, TA100	Ames test. Tested in a preincubation assay. Test concentrations unspecified.	Mutation	With and without	-ve  A limited assay. Only 2 strains were tested, however current protocols recommend using at least 4 strains.	Zeiger & Margolin, 2000
<i>Salmonella typhimurium</i> , strains TA98, TA100, TA1535, TA1537, TA1538	Ames test. Tested at concentrations of up to 10 mg/plate.	Mutation	With and without	-ve	Bonin & Baker, 1980
<i>Salmonella typhimurium</i> , various strains including TA98, TA100, TA1535, TA1537, TA1538	3 Ames tests cited in a JECFA review. Limited details available from the expert review.	Mutation	With and without	-ve	Brusick, 1976; Muzzall & Cook, 1979; Viola & Nosotti, 1978
<i>Salmonella typhimurium</i> , various strains	Various other Ames tests.  The full papers for these studies were not obtained as the results are in line with the other Ames test	Mutation	With and without	-ve	Brown & Dietrich, 1983; Chung et al. 1981; Ozaki et al. 1998; Prival et al. 1988; Rafii et

	reports consulted, which include high quality studies.				al. 1997
<i>Salmonella typhimurium</i> , strains TA1535, TA1537 and/or TA98, TA100	The effect of azo-reduction on the mutagenic potential of allura red was examined. Allura red was preincubated (aerobically and anaerobically), or treated with Clostridium bacteria (which have azoreductase activity), or flavin mononucleotide (a facilitator of azo reduction), before being tested in the Ames assay.	Mutation	With and without	-ve	Brown & Dietrich, 1983; Brown et al. 1978; Prival et al. 1988; Rafii et al. 1997
<i>Salmonella typhimurium</i> , strain TA1538	Liquid fluctuation test. Cells were treated with a concentration of 10 mg/ml for 72-96 hr and the increase in turbidity was assessed.	Mutation	With and without	-ve  A limited assay as only tested in one strain.	Haveland-Smith & Combes, 1980
<i>Escherichia coli</i> , strain WP2uvrA	Liquid fluctuation test. Cells were treated with a concentration of 10 mg/ml for 72-96 hr and the increase in	Mutation	With and without	-ve	Haveland-Smith & Combes, 1980

	turbidity assessed.				
<i>Escherichia coli</i> , wild-type strain (WP2trp uvr A), and two strains deficient in DNA repair capability (WP67 trp uvr A pol A and WP100 trp uvr A rec A)	<p>Differential killing assays (rec and pol assays). Cells treated with a concentration of 10 mg/ml for 3.5 hr (repair-proficient cells (WP2); both rec and pol assays), 5.5 hr (repair-deficient cells (WP100); rec assay) and 4.5 hr (repair-deficient cells (WP67); pol assay).</p> <p>DNA damage is indicated by a higher toxicity in the DNA repair deficient strains compared with the wild-type strains.</p>	<p>DNA damage</p> <p>(indicative test)</p>	With and without	-ve	Haveland-Smith & Combes, 1980
<i>Escherichia coli</i> , two wild-type strains (AB1157, CSH7) and three strains deficient in DNA repair capability (TN1005, RPC501, UM1)	<p>Differential killing assays. Treated with allura red and illuminated with visible light for 18 hr. Concentrations not specified.</p> <p>DNA damage is indicated by a difference in toxicity between the wild and DNA repair</p>	<p>DNA damage</p> <p>(indicative test)</p>	Without	<p>-ve</p> <p>A limited study as no activation was used.</p>	Kuraoka et al. 1991

	deficient strains.				
Human, MCF-7 breast cancer cells	Cells treated with allura red and assayed for p53-DNA binding.  No further details available.	DNA damage	Not specified	-ve	Dees et al. 1997
<i>Saccharomyces cerevisiae</i> , 3 strains	Genetic tests.  No further details of this unpublished study were available from the expert review.	Chromosome damage	With and without	-ve	Anon, 1977b
<i>Saccharomyces cerevisiae</i> , 1 strain	Mutation assay.  No further details of this unpublished study were available from the expert review.	Mutation	Probably with and without	-ve	Brusick, 1976

## References

Anon (1977a). Mutagenicity investigations with allura red. Sex-linked recessive lethals, *Drosophila melanogaster*, Bowling Green University, Ohio, USA. Interim report of the Working Group on FD & C Red no. 40, 19 January 1977, submitted to WHO by the US Food and Drug Administration (cited in JECFA, 1980).

Anon (1977b). Mutagenicity investigations with allura red. *Saccharomyces cerevisiae*. Genetic Toxicology Branch of the FDA. Interim report of the Working Group on FD & C Red no. 40, 19 January 1977 (cited in JECFA, 1980).

Anon (1978). Evaluation of substances of interest for genetic damage using *Drosophila melanogaster*. Unpublished report submitted to WHO by the United States Food and Drug Administration, 31 March 1978 (cited in JECFA, 1980).

Bonin A M & Baker R S U (1980). Mutagenicity testing of some approved food additives with the Salmonella/microsome assay. Food Technology in Australia 32, 608-611.

Brown J P & Dietrich P S (1983). Mutagenicity of selected sulfonated azo dyes in the Salmonella microsome assay: use of aerobic and anaerobic activation procedures. Mutation Research 116, 305-316 (cited in Toxline).

Brown J P et al. (1978). Mutagenicity testing of certified food colors and related azo, xanthene and triphenylmethane dyes with the Salmonella/microsome system. Mutation Research 56, 249-272.

Brusick D (1976). Mutagenicity evaluation NTRZ-4576 (Allura Red AC). Unpublished report number 2547 by Litton Bionetics, Inc., submitted to WHO by Allied Chemical Corporation (cited in JECFA, 1980).

CCRIS (2001). FD&C red no. 40. CCRIS record number 3493. Chemical Carcinogenesis Research Information System.

Chung K-T et al. (1981). Mutagenicity testing of some commonly used dyes. Applied and Environmental Microbiology 42, 641-648 (cited in Toxline).

Dees C et al. (1997). Estrogenic and DNA-damaging activity of Red No. 3 in human breast cancer cells. Environmental Health Perspectives 105 (Supplement 3), 625-632.

Haveland-Smith R B & Combes R D (1980). Screening of food dyes for genotoxic activity. Food and Cosmetics Toxicology 18, 215-221.

JECFA (1980). Toxicological evaluation of certain food additives. Twenty-fourth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Food Additives Series 15. World Health Organization, Geneva.

Jorgenson T A et al. (1978). Study of the mutagenic effects of FD & C Red 40 (75-60) by the heritable translocation study in mice. Unpublished report LSU-5588 by SRI International, supplied by the United States Food and Drug Administration (cited in JECFA, 1980).

Kawaguchi S et al. (2001). Evaluation of in vivo genotoxicity of twelve synthetic tar dyes permitted in Japan using mouse comet assay. Mutation Research 483 (Supplement 1), S170.

Kuraoka I et al. (1991). Sensitivity of E. coli mutants lacking active oxygen defense systems to photodynamic action. Mutation Research 253, 260.

Lee W R et al. (1983). The sex-linked recessive lethal test for mutagenesis in Drosophila melanogaster. A report of the U.S. Environmental Protection Agency Gene-Tox Program. Mutation Research 123, 183-279.



Muzzall J & Cook W (1979). Mutagenicity test of dyes used in cosmetics with Salmonella/mammalian-microsome test. Mutation Research 67, 1-8 (cited in JECFA, 1980).

NTP [undated]. Salmonella study overview: allura red C.I. 16035. Study ID A81251 ([http://ntp-apps.niehs.nih.gov/ntp\\_tox/index.cfm](http://ntp-apps.niehs.nih.gov/ntp_tox/index.cfm); search using CAS no. 25956-17-6).

Ozaki A et al. (1998). Mutagenicity and DNA-damaging activity of decomposed products of food colours under UV irradiation. Food and Chemical Toxicology 36, 811-817 (cited in CCRIS, 2001).

Prival M J et al. (1988). Evaluation of azo food dyes for mutagenicity and inhibition of mutagenicity by methods using Salmonella typhimurium. Mutation Research 206, 247-259 (cited in Toxline).

Rafii F et al. (1997). Mutagenicity of azo dyes used in foods, drugs and cosmetics before and after reduction by Clostridium species from the human intestinal tract. Food and Chemical Toxicology 35, 897-901 (cited in CCRIS, 2001).

Sasaki Y F et al. (2002a). The comet assay with 8 mouse organs: results with 39 currently used food additives. Mutation Research 519, 103-119.

Sasaki Y F et al. (2002b). The comet assay with eight mouse organs: results with currently used food dyes by single and multiple treatment. Mutagenesis 17, 572.

Tsuda S et al. (2001). DNA damage induced by red food dyes orally administered to pregnant and male mice. Toxicological Sciences 61, 92-99.

Viola M & Nosotti A (1978). Application of the Ames test to certain color additives. Bollettino Chimico Farmaceutico 117, 402-415 (cited in JECFA, 1980).

Zeiger E & Margolin B H (2000). The proportions of mutagens among chemicals in commerce. Regulatory Toxicology and Pharmacology 32, 219-225.